

LUTEOLIN 7,4'-DIMETHYL ETHER 3'-GLUCOSIDE FROM *GELONIUM MULTIFLORUM*

NAZNEEN PARVEEN and NIZAM U. KHAN

Department of Chemistry, Aligarh Muslim University, Aligarh 202 001, India

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Key Word Index—*Gelonium multiflorum*; Euphorbiaceae; luteolin 7,4'-dimethyl ether 3'-glucoside.

Abstract—A new flavone glycoside has been isolated from the leaves of *Gelonium multiflorum* and characterized as luteolin 7,4'-dimethyl ether 3'-glucoside.

INTRODUCTION

Gelonium, a genus of shrubs and small trees is distributed in the tropical and sub-tropical parts of Asia and Africa. Three species of *Gelonium* from India are well known for their medicinal properties [1]. We now report the characterization of a new flavone glycoside (1) from leaves of *G. multiflorum*.

RESULTS AND DISCUSSION

Compound 1 analysed for $C_{23}H_{24}O_{11}$, responded to the Shinoda test [2], gave a positive Molisch test and reduced Tollen's reagent. The UV spectrum showed λ_{max} at 285 and 335 nm, and the IR spectrum exhibited absorption bands due to hydroxyl (3400 cm^{-1}), an α,β -unsaturated ketone ($1650, 1601\text{ cm}^{-1}$) and an aromatic ring ($900, 850, 770\text{ cm}^{-1}$). The characteristic colour reactions and spectral properties indicated 1 to be a flavone glycoside.

Hydrolysis of 1 with both 6% HCl and β -glucosidase yielded D-glucose and an aglycone (2) which was identical with an authentic sample of luteolin 7,4'-dimethyl ether (pillonin) [3, 4]. The presence of a free hydroxyl at C-5 [5] was confirmed by a bathochromic shift with aluminium chloride. The formation of a diacetate and a tetramethyl ether supported the presence of two free hydroxyls and two free methoxyl groups in 2.

The $^1\text{H NMR}$ spectrum of 1 in $\text{DMSO}-d_6$ showed an AMX system due to a 3',4'-disubstitution. An *ortho* coupled doublet of one proton at $\delta 7.68$ ($J = 8\text{ Hz}$), a double doublet of one proton at $\delta 7.85$ ($J = 2$ and 8 Hz) and a *meta* coupled doublet of one proton at $\delta 7.63$ ($J = 2\text{ Hz}$) are assigned to H-5', H-6', and H-2', respectively. The 5,7 substitution pattern was demonstrated by the presence of two *meta* coupled doublets of one proton each at $\delta 6.60$ and $\delta 7.03$, ascribable to H-6 and H-8, respectively. A sharp singlet at $\delta 6.90$ is assigned to the H-3 proton. Two singlets at $\delta 3.87$ and $\delta 3.90$ integrating for three protons each, confirmed the presence of two aromatic methoxyls. The anomeric proton, H-1" of the glucose appeared as a doublet at $\delta 5.69$ ($J = 7\text{ Hz}$). The chemical shift confirmed the direct attachment to the aglycone and the diaxial coupling ($J = 7\text{ Hz}$) between

H-1" and H-2" suggested a β -configuration. These data indicated that 1 is a 5,7,3',4'-oxygenated compound and the mass spectrum gave a base peak of $[\text{M} - \text{glucose}]^+$ at m/z 314.

The position of the linkage of glucose was established by comparing the UV and $^1\text{H NMR}$ spectra of the glycoside (1) and the aglycone (2). Since 2 has free 5 and 3' hydroxyls the glucose in 1 must be attached to one of these positions. A bathochromic shift of 20 nm with aluminium chloride leaves only the 3' position free for glycosylation. This was further confirmed by methylation and subsequent hydrolysis of the methylated product with 6% HCl to give a partial methyl ether characterized as 5,7,4'-trimethoxyluteolin by spectral studies. The sugar was confirmed as glucose by co-paper chromatography with an authentic sample and periodate oxidation indicated that it was in the pyranose form [6]. Compound 1 is therefore identified as luteolin 7,4'-dimethyl ether 3'-O- β -D-glucoside.

EXPERIMENTAL

All mps are uncorr., UV and IR spectra were recorded in MeOH and as KBr pellets, $^1\text{H NMR}$ were measured at 60 MHz in CDCl_3 with TMS as int. standard.

Extraction and isolation. Air dried leaves of *G. multiflorum* A. Juss. collected from FRI, Dehradun, were refluxed with MeOH. The MeOH concentrate was successively treated with petrol (60–80°), C_6H_6 and EtOAc. The EtOAc concentrate was chromatographed over silica gel. Elution with EtOAc afforded 1 mp $> 300^\circ$, which crystallized from EtOAc-Me₂CO as pale yellow needles. It gave a positive colour with Mg-HCl (orange) and FeCl_3 (green). $^1\text{H NMR}$ (60 MHz, $\text{DMSO}-d_6$): δ 6.60, 7.03 (1H each, d , $J = 2\text{ Hz}$, H-6 and H-8), 6.90 (1H, s , H-3), 7.68 (1H, d , $J = 8\text{ Hz}$, H-5'), 7.85 (1H, dd , $J = 2$ and 8 Hz , H-6'), 7.63 (1H, d , $J = 2\text{ Hz}$, H-2'), 3.87, 3.90 (3H each, OCH_3 -7, 4'), 3.30–4.0 (6 glucosyl protons). UV $\lambda_{max}^{\text{MeOH}}$ nm: 285, 335; + AlCl_3 : 296, 355; + AlCl_3/HCl : 296, 355; + NaOAc: 288, 335; + NaOAc/ H_3BO_3 : 288, 335; + NaOMe: 290, 335. MS m/z : 314 $[\text{M} - \text{glucose}]^+$, 285 $[\text{M} - 29]^+$, 271 $[\text{M} - 43]^+$, 268 $[\text{M} - 46]^+$, 167 $[\text{C}_8\text{H}_7\text{O}_4]$, 148 $[\text{C}_9\text{H}_8\text{O}_2]$, 138 $[\text{C}_7\text{H}_6\text{O}_3]$, 133 $[\text{C}_8\text{H}_5\text{O}_2]$ and 123 $[\text{C}_7\text{H}_7\text{O}_2]$.

Acid hydrolysis of 1 with 6% HCl at 100° for 2 hr gave yellow crystals of an aglycone (2), mp 236°. When analysed for $C_{17}H_{12}O_{16}$, $[M]^+ = 314$ it was identical with authentic luteolin 7,6'-dimethyl ether (mp, MS, UV and IR). The acetate of 2, mp 185°, was also identical (MS and NMR) with authentic sample [3]. Compound 1 was also methylated and then hydrolysed with 6% HCl to give 5,7,4'-trimethoxy luteolin, mp 220°, UV λ_{max}^{MeOH} nm: 285, 335; + $AlCl_3$: 287, 335; + $AlCl_3/HCl$ 287, 335; + NaOAc 285, 335; + NaOAc/ H_3BO_3 285, 335; + NaOMe 290, 335. The identity of the sugar was confirmed as glucose by Co-PC with an authentic marker in *n*-butanol-HOAc-H₂O using aniline hydrogen phthalate as detection reagent.

REFERENCES

1. Sastri, B. N. ed. (1956) *The Wealth of India*, CSIR, New Delhi, p. 122.
2. Shinoda, J. (1928) *J. Chem. Pharm. Soc. Japan* 48, 214.
3. Nunez-Alarcon, J. (1971) *J. Org. Chem.* 36, 2829.
4. Jain, A. C. and Sharma, B. N. (1973) *Phytochemistry* 12, 1455.
5. Markham, K. R. and Mabry, T. J. (1975) in *The Flavonoids* (Harborne, J. B., Mabry, T. J. and Mabry, H., eds), Chapman and Hall, London.
6. Sinha, K. S., Sinha, S. K. and Dewivedi, N. (1985) *J. Ind. Chem. Soc.* LXII, 169.

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FLAVONOID AGLYCONES FROM *XANTHOCEPHALUM GYMNOSPERMOIDES* VAR. *GYMNOSPERMOIDES*

SANGGONG YU,* NIANBAI FANG† and TOM J. MABRY

The University of Texas at Austin, Austin, TX 78713, U.S.A.

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Key Word Index—*Xanthocephalum gymnospermoides* var. *gymnospermoides*; Compositae; Astereae; 5,7,4'-trihydroxy-3,3',5'-trimethoxyflavone.

Abstract—Six known flavonoid aglycones and the newly described 5,7,4'-trihydroxy-3,3',5'-trimethoxyflavone were isolated from *Xanthocephalum gymnospermoides* var. *gymnospermoides*.

INTRODUCTION

In 1961, Solbrig recognised eight species for *Xanthocephalum* [1], while Lane (1980) treated the genus as having five species with other taxa being transferred to *Gutierrezia* [2]. As part of our continuing chemical systematics investigation of the 'Gutierrezia-*Xanthocephalum* complex' [3–9], we report here the isolation of seven flavonoids from the aerial parts of *Xanthocephalum gymnospermoides* (Gray) B. and H. var. *gymnospermoides*.

RESULTS AND DISCUSSION

Column chromatography and preparative paper chromatography of the material from the dichloromethane and ethyl acetate extracts of a concentrated aqueous methanol extract of the aerial parts of *X. gymnospermoides* var. *gymnospermoides* afforded flavonoids 1–7 including one new compound. The new subst-

ance is 5,7,4'-trihydroxy-3,3',5'-trimethoxyflavone (1) and the six known compounds are 5,3',5'-trihydroxy-3,7,4'-trimethoxyflavone (2), 5,7,4'-trihydroxy-3',5'-dimethoxyflavone (3), 5,7,4'-trihydroxy-3,3'-dimethoxyflavone (4), 5,3',4'-trihydroxy-3,7-dimethoxyflavone (5), 3,5,7,3',4'-pentahydroxyflavone (6) and 5,7,4'-trihydroxy-3-methoxyflavone (7). The flavonoid chemistry of this taxon is different from that of woody species of *Gutierrezia* [3–6] and the related monotypic *Gymnosperma glutinosum* [9]; these latter taxa produce large quantities of flavonoids with 6,8-oxygenation as well as flavonol 3-methyl ethers with rare 2'-oxygenation. In contrast, the flavonoids isolated from *Xanthocephalum gymnospermoides* var. *gymnospermoides* are similar to those of two herbaceous species of *Gutierrezia*, *G. wrightii* [8] and *G. alamanii* var. *megalocephala* [7], two taxa formerly placed in *Xanthocephalum* [2]. The species investigated here and the latter two species all yielded only a few flavonoids, which all lacked 6,8-oxygenation and 2'-oxygenation. Detailed data are presented only for compound 1.

¹H NMR spectrum in carbon tetrachloride of the TMSi ether of 1 showed one two-proton singlet which was assigned to protons at 2' and 6' (δ 7.30 ppm) in a symmetrically substituted B-ring and two one-proton doublets at δ 6.15 ($J = 2.5$ Hz) and 6.48 ($J = 2.5$ Hz) characteristic of H-6 and H-8, respectively [10]. The remaining signals in

*Permanent address: Wuhan Institute of Medical Sciences, Wuhan, China.

†Permanent address: Hubei College of Chinese Traditional Medicine, Wuhan, China.